

## Review

## Matrix Metalloproteinase-8 and Cardiovascular Diseases: A Narrative Review

<sup>1</sup>Lütfü Aşkın , <sup>2</sup>Hüsna Şengül Aşkın , <sup>3</sup>Okan Tanrıverdi 

<sup>1</sup>Department of Cardiology, Gaziantep Islamic Science and Technology University, Gaziantep, Türkiye

<sup>2</sup>Department of Infectious Disease, Gaziantep City Hospital, Gaziantep, Türkiye

<sup>3</sup>Department of Cardiology, Adiyaman Education and Research Hospital, Adiyaman, Türkiye

**Corresponding Author:** Lütfü Aşkın, M.D., Department of Cardiology, Gaziantep Islamic Science and Technology University, Gaziantep, Türkiye. **E-mail:** lutfuaskin23@gmail.com

Submitted at: 09.10.2024 - Accepted at: 12.12.2024 - Published at: 30.12.2024  
The journal is licensed under: Attribution 4.0 International (CC BY 4.0)

Avicenna Anatol J Med. Year; 2024, Volume: 1, Issue: 1

 [10.5281/zenodo.14568908](https://doi.org/10.5281/zenodo.14568908)

## Abstract

Matrix metalloproteinases-8 (MMP-8), commonly known as neutrophil collagenase, is an MMP enzyme. MMPs break down extracellular matrix components, which support tissues. Skin, bones, and blood vessels contain collagen, which MMP-8 degrades. Researchers have linked higher MMP-8 levels to atherosclerosis, heart attacks, and heart failure (HF). MMP-8 damages the extracellular matrix in heart tissue and blood vessels, which can worsen cardiovascular diseases (CVDs) by altering the shape of artery walls, destabilizing plaques, and changing the shape of ventricles. MMP-8 breaks down collagen in the fibrous cap of atherosclerotic plaques, weakening it and increasing the likelihood of plaque rupture, which may lead to a heart attack or stroke. MMP-8 activity accelerates ventricular remodeling in heart failure, impairing cardiac function. MMP-8's role in CVDs is complex. Certain research suggests that MMP-8 may assist in the repair of injured tissue. How MMP-8 activity impacts CVDs overall depends on the efficiency of MMP-8 and its tissue inhibitors, or TIMPs, working together. MMP-8 inhibitors may reduce matrix breakdown and tissue damage in CVDs. MMP-8's importance in CVDs and its therapeutic potential requires further investigation. A trained healthcare professional should diagnose and treat CVDs and other medical conditions.

**Keywords:** Atherosclerosis, Cardiovascular diseases, Matrix metalloproteinases-8

## INTRODUCTION

Matrix metalloproteinases (MMPs) are a family of enzymes that play a role in the remodeling and degradation of the extracellular matrix (ECM), which is the structural framework of tissues and organs in the body. Secreted as inactive enzymes (proMMPs), MMPs activate in response to various physiological and pathological stimuli. MMPs are involved in many physiological processes, such as embryogenesis, tissue repair, and tissue remodeling during normal development and growth. Pathological conditions such as inflammation, tissue injury, and cancer metastasis also implicate MMPs. MMPs are capable of degrading components of the ECM, including collagen, elastin, and proteoglycans, which are essential for maintaining tissue structure and function (1).

Fibroblasts, macrophages, neutrophils, and endothelial cells are just a few of the various cell types that produce and secrete MMPs. A complex interplay of activators and inhibitors tightly regulates their activity to prevent excessive tissue remodeling. Many diseases, including

arthritis, tissue fibrosis, cardiovascular diseases (CVDs), and cancer metastasis, have implicated MMP dysregulation. Researchers have developed MMP inhibitors as potential therapeutic agents for these diseases, but the complex roles of MMPs in tissue remodeling and their potential side effects have limited their clinical use. Overall, MMPs are important enzymes involved in tissue remodeling and play a crucial role in both physiological and pathological processes in the body (1).

Matrix metalloproteinase-8 (MMP-8), also known as neutrophil collagenase, is a member of the matrix metalloproteinase family that specifically degrades collagen, which is a major component of the ECM in tissues. White blood cells known as neutrophils, which play a role in the immune response to infections, are the main producers of MMP-8. In the context of cardiovascular disease, MMP-8 has been implicated in the pathogenesis of atherosclerosis, which is a condition characterized by the accumulation of plaques in the

arterial walls. Atherosclerosis is a major risk factor for CVDs such as coronary artery disease, stroke, and peripheral artery disease. It is believed that MMP-8 contributes to atherosclerosis by breaking down the collagen fibers in the arterial walls, weakening the ECM's structural integrity and potentially causing plaque formation. Researchers have found that atherosclerotic lesions upregulate MMP-8, especially in the areas where plaques rupture, releasing their contents into the bloodstream and potentially triggering acute cardiovascular events (2).

The balance between MMPs and their tissue inhibitors (TIMPs) is critical for maintaining ECM homeostasis. If this balance is out of whack, with more MMP activity and/or lower TIMP levels, the ECM can break down too quickly, which can help heart diseases like atherosclerosis start and get worse. Several studies have suggested that MMP-8 may be a potential therapeutic target for CVDs. Preclinical and clinical studies have investigated MMP-8 inhibitors, but we are still evaluating their efficacy and safety in human subjects. Additionally, researchers are exploring other strategies as potential therapeutic approaches, such as modulating the activity of TIMPs or targeting other MMPs involved in CVDs. This review updates data suggesting that the neutrophilic collagenase MMP-8 may directly cause CVDs.

MMP-8 protein or mRNA is expressed higher in symptomatic unstable carotid plaques, abdominal aortic aneurysm aortas, rupture sites, and advanced shoulder atherosclerosis lesions. These findings implicate MMP-8 in matrix remodelling in atherosclerosis and unstable plaque rupture—the pathophysiology of acute coronary syndromes. It is rare to examine CVD serum or plasma MMP-8 levels, especially in a prospective manner. Plasma MMP-8 affects coronary artery disease, carotid plaque instability, morphology, and stroke delay. Congestive cardiac failure and cerebral ischaemia reduce plasma MMP-8. Small sample sizes and low statistical power may cause these disparities. Thus, the study's sample size, prospective population-based design, and extended follow-up are positives. A few CVD fatalities were analyzed. Solid outcomes. The male sample necessitates further investigation into MMP-8's role in women's CVD.

## OVERVIEW

MMP-8 protein or mRNA is expressed higher in symptomatic unstable carotid plaques, abdominal aortic aneurysm aortas, rupture sites, and advanced shoulder atherosclerosis lesions. These findings implicate MMP-8 in matrix remodelling in atherosclerosis and unstable plaque rupture—the pathophysiology of acute coronary syndromes. It is rare to examine CVD serum or plasma MMP-8 levels, especially in a prospective manner. Plasma MMP-8 affects coronary artery disease, carotid

plaque instability, morphology, and stroke delay. Congestive cardiac failure and cerebral ischaemia reduce plasma MMP-8. Small sample sizes and low statistical power may cause these disparities. Thus, the study's sample size, prospective population-based design, and extended follow-up are positives. A few CVD fatalities were analyzed. Solid outcomes. Due to the male sample, MMP-8's role in women's CVD needs more investigation (3,4).

Studies show subclinical atherosclerosis with carotid artery intima media thickness (IMT) above 1 mm. IMT increases stroke and MI. Males with IMT >1 mm exhibited multiple CVD risk factors, including elevated fibrinogen. Plasma fibrinogen concentrations significantly and positively correlated with serum MMP-8 and the MMP-8/tissue inhibitor of metalloproteinase-1 (TIMP-1) ratio after adjusting for key CVD risk variables. Inflammation raises fibrinogen, causing CVDs. A high fibrinogen concentration, with a multivariate RR of 1.65 (1.05 to 2.60,  $p=0.029$ ), predicted AMI in men without preclinical atherosclerosis, but not other endpoints or subclinical conditions. Smoking altered MMP-8 and fibrinogen levels. Smokers had slightly greater serum MMP-8 than nonsmokers, although urine nicotine metabolites and smoking years were unrelated (5,6).

Preclinical atherosclerosis may increase serum MMP-8. Periodontal bacteria inflaming tissues may produce MMP-8 in the blood. Researchers recently linked periodontitis severity and seropositivity to subclinical atherosclerosis. The combined serum antibody level to major pathogens is used as a confounding factor in the linear regression analysis. This is because periodontitis and seropositivity to periodontal pathogens are two different CVD risk factors. Antibody and MMP-8 levels were unrelated. In basic investigations, there was no significant difference in antibody levels to periodontal pathogens between subclinical atherosclerosis and IMT 1 mm. Blocking MMPs and TIMPs prevents plaque rupture (7). Widespread or subclinical atherosclerosis was associated with a worse cardiovascular prognosis and higher serum MMP-8 levels (8).

## CORONARY ARTERY DISEASE

MMPs regulate collagen production and breakdown, which impacts plaque development, susceptibility, and vascular remodelling. Only MMP-1, MMP-8, and MMP-13 break collagen, leaving it vulnerable to further degradation by MMP-2 and MMP-9. MMP-8 destroys type I collagen, a key component of atherosclerotic plaques, 3-fold faster than MMP-1 and MMP-13. Atherosclerotic plaques upregulate MMP-1 and MMP-13, while neutrophils, which are rare in plaques, exclusively generate MMP-8. Endothelial cells, smooth muscle cells, and macrophages in human atherosclerotic plaques make MMP-8 mRNA and protein. These cells

are found in lipid-rich plaques with a thin fibrous cap. MMP-8 breaks down atherosclerotic plaque collagen, causing plaque vulnerability or vascular remodelling. We have not yet established a link between MMP-8 and coronary artery disease (CAD). Increased plasma MMP-8 levels independently correlate with CAD. MMP-8 concentration correlates with CAD severity, peaking in patients with 3-vessel disease. MMP-8 concentration independently causes three-vessel disease. High MMP-8 levels in CAD patients may target the adaptive remodeling of coronary arteries or plaque vulnerability (9).

Plaque vulnerability predicts unstable angina and myocardial infarction (MI) in angiography-detected complicated coronary lesions. Complex lesions are common in unstable angina and 10–20% in stable angina. Even after eliminating acute MI and unstable angina, complex lesions were present in 23% of CAD patients. Serum MMP-1 values in 185 stable CAD patients were similar, but high quantities were associated with complex coronary lesions. MMP-8 levels were higher in complex lesions, although not statistically significant. Plasma MMP-8 in stable CAD patients may indicate coronary atherosclerosis severity rather than plaque susceptibility (Figure 1). [10]. Coronary events were more common in high-hsCRP CAD patients. Independent of HsCRP levels, MMP-8 levels predicted CAD. Vascular MMP-9 activates diabetes [11]. High hyperglycemia in vascular cells enhanced MMP-1, MMP-2, and MMP-9 activity, according to Death et al. [12] Independent of diabetes, we linked high plasma MMP-8 levels to CAD, particularly 3-vessel disease. Statins lower rabbit atheroma MMP-1, MMP-3, and MMP-9 expression and CAD plasma MMP-9.

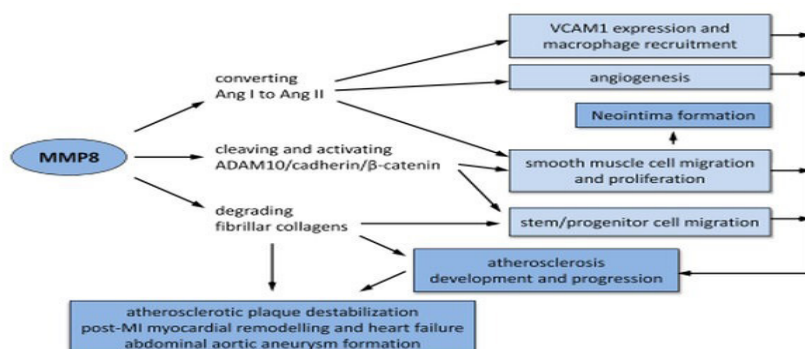
## PULMONARY ARTERIAL HYPERTENSION

The lung upregulates MMP-8 after monocrotaline (MCT), but its role in pulmonary arterial hypertension (PAH) is uncertain. Severe PAH patients and three animal PAH models had elevated plasma MMP-8 (13). pulmonary vascular MMP-8 expression is linked to

the severity of PAH, the development of PAH in mice exposed to hypoxia, and changes in the blood vessels. Mice without MMP-8 develop chronic hypoxia-induced PAH, vascular remodelling, and RV dysfunction. PAH development seems to include adaptive MMP-8 overexpression. Hypoxaemia, arrhythmia, and cardiac ischaemia may potentially cause mortality (14).

The activity of elastases and MMPs may directly improve the mitogenic activity of pulmonary artery smooth muscle cells (PASMCs) by releasing mitogens, revealing hidden integrin binding sites, or turning on mitogen receptors. MMP activity activates latent integrin-b3 binding sites by upregulating tenascin-C synthesis and deposition. Integrin-interacting matricellular proteins induce PAH. Osteopontin-expressing PAH patients had better outcomes (15,16). PASMCs and adventitial fibroblasts proliferate and undergo mitogenesis (17,18). Human PAH elevates thrombospondin-1, which correlates with severity and cardiac output, and significantly expresses in end-stage disease distal PAs (19-22). Thrombospondin-1 and integrin-b3 may increase PASMC migration and proliferation (23).

MMPs control integrin signaling. Gross MMP inhibition in cancer cell lines lowers traction pressures, cell rounding, and cell softness through integrin-B1 levels and localization (24). MMP-mediated integrin-b3 activation revealed no collagen-cleaved integrin binding sites. TIMP-1 activates integrin-b1 through CD63, whereas MMP-8 suppresses it in cancer cells (25-27). These results complement our observation that MMP-8 depletion enhances integrin signaling via integrin B3, not B1. Integrin-b3, crucial for shape signal transduction in SMCs, plays a role in SMC mobility, including chemotaxis (28,29). These signals are sent by Merlin, a Yes-associated protein/transcriptional co-activator with a PDZ binding motif (YAP/TAZ) regulator (30). PAH development increases pulmonary vasculature MMP-8, a new protective factor. MMP-8 disrupts PASMC integrin-β3/FAK, YAP/TAZ-dependent mechanical signaling, and matrix composition to resist pathologic mechanobiological feedback (31).



**Figure 1.** MMP8 causes atherosclerosis, plaque destabilisation, and myocardial remodelling. Mechanism diagrams. Other publication containing the figure in the manuscript include "Pharmacol Ther 2015;147:111-22". MMP8: Matrix Metalloproteinase-8, Ang I: Angiotensin I, Ang II: Angiotensin II, ADAM10: A Disintegrin And Metalloproteinase 10, VCAM1: Vascular Cell Adhesion Molecule 1, MI: Myocardial Infarction

## ATRIAL FIBRILLATION

Atrial fibrillation (AF) is a common cardiac arrhythmia characterized by rapid and irregular electrical activity in the atria, or upper chambers, of the heart. It is associated with an increased risk of stroke, heart failure, and other cardiovascular complications. Emerging evidence suggests that MMP-8, a member of the matrix metalloproteinase family, may be involved in the pathogenesis of atrial fibrillation. MMP-8 is a white blood cell type that is primarily responsible for producing the enzyme, and it participates in tissue remodelling by destroying the ECM. MMP-8 is associated with the structural remodelling of the atrial tissue, a hallmark of atrial fibrillation. The atrial ECM changes during structural remodelling. These changes include fibrosis (too much collagen buildup) and dilation (enlargement of the atria). These can stop the heart's normal electrical conduction and help AF start and stay active.

Researchers have found that patients with AF have upregulated MMP-8 in their atrial tissue, and they have correlated its expression with the severity of atrial fibrosis. Studies have also demonstrated that MMP-8 stimulates the breakdown of collagen in the atrial extracellular matrix (ECM), thereby exacerbating the atrial tissue's structural integrity and accelerating the onset of atrial fibrillation (AF). Apart from its involvement in structural remodelling, MMP-8 also plays a role in inflammation, oxidative stress, and the immune response, all of which can contribute to the pathophysiology of AF. MMP-8 can modulate the activity of other inflammatory molecules and enzymes, and its increased expression may lead to an imbalance in the inflammatory response in the atrial tissue promoting AF development. Targeting MMP-8 has emerged as a potential therapeutic strategy for AF, despite the incomplete understanding of the exact mechanisms through which MMP-8 contributes to AF pathogenesis and the need for further research. Preclinical studies have investigated MMP inhibitors, including specific inhibitors of MMP-8, and have shown promise in reducing atrial fibrosis and AF vulnerability in animal models. However, limited clinical studies in humans have yet to establish the safety and efficacy of MMP inhibitors for AF treatment.

## ISCHEMIC STROKE

The pathophysiology of stroke has implicated MMP-8. Research has demonstrated an increase in MMP-8 levels in the brain tissue and blood of stroke victims. Studies have demonstrated that MMP-8 degrades the extracellular matrix in the brain, including the basement membrane of blood vessels, potentially leading to disruption of the blood-brain barrier and inflammation. MMP-8 can also activate other MMPs, leading to a cascade of matrix degradation and tissue damage. Animal models have demonstrated the role of MMP-

8 in stroke progression, where a deficiency in MMP-8 leads to reduced brain injury and improved outcomes. Researchers have investigated MMP-8 inhibitors as potential therapeutic targets for stroke to reduce tissue damage and inflammation (32).

MMPs damage ischaemic stroke brains. Stroke patients' infarcted brains express higher MMP-1, -2, -3, -8, -9, -10, -13, and TIMP-1 than non-ischaemic areas (33). Microglia, macrophages, and neutrophils produce blood-brain barrier leakage and neurovascular abnormalities. Stroke therapy targets MMPs. We must carefully design treatment suppression because MMPs are essential for angiogenesis during recovery. MMP-9 is the most studied MMP in CVD; however, other MMP family members may have therapy potential. Stroke patients seldom study MMP-8 (34).

## RECENT STUDIES

Neuropeptide Y upregulates MMP-8, which increases macrophage migration and neointima development (35). Iwańczyk et al. linked MMP-8 to aberrant coronary artery dilation (36). Zhang et al. found increased MMP-8 levels in aortic dissection patients (37). Having the MMP-8 promoter GTTT haplotype lowers the production of MMP-8 during inflammation and raises the risk of high blood pressure (38,39). Researchers have linked low blood levels of MMP-12 to ischemic stroke, low levels of MMP-1 and MMP-12 to large-artery stroke, and high levels of MMP-8 to lacunar stroke. cunar stroke (38,39). In their study, Yang et al. found that MMP8 from macrophages changes how AdSPC turns into SMC and how neointimal SMC grows after an injury (40). The study of MMP8's part in AdSPC differentiation and neointima development in angiographic restenosis could lead to the creation of new drugs that can stop this condition.

An imbalance in TIMP-MMP changes the structure of the extracellular matrix (ECM) and leads to the remodelling of dilated cardiomyopathy (DCM). This process could help researchers come up with new ways to treat DCM (41). MSSA pneumonia has implicated neutrophil proteases, including MMPs, in a rare aortic rupture (42). **Table 1** displays the main topics of recent studies.

## CONCLUSION

Neutrophil collagenase (MMP-8) is an MMP enzyme. MMPs degrade the extracellular matrix, which supports tissues. MMP-8 breaks down collagen in skin, bones, and blood vessels. Higher MMP-8 levels are associated with atherosclerosis, heart attacks, and heart failure. MMP-8 breaks down the extracellular matrix in heart tissue and blood vessels. This changes the shape of the artery walls, makes plaques less stable, and changes the shape of the ventricles, all of which can lead to CVDs. MMP-8 destroys collagen in atherosclerotic plaques'

**Table 1.** The main topic points of recent studies.

Reference no.	Authors	Subjects	Main theme
Ref (16)	Mura et al.	Pulmonary arterial hypertension	Osteopontin (OPN) expression is closely linked with disease severity in severe PAH patients' lungs. PAH vascular remodelling may include OPN.
Ref (19)	Kumar et al.	Pulmonary arterial hypertension	TGF- $\beta$ -dependent vascular disorders may be treated by targeting thrombospondin-1-dependent TGF- $\beta$ activation.
Ref (22)	Rogers et al.	Pulmonary arterial hypertension	CD47 targets cMyc to boost endothelin-1 signalling in pre-clinical PH models. Clinical PH upregulates TSP1-CD47, which causes pulmonary arterial vasculopathy and dysfunction.
Ref (23)	Rogers et al.	Pulmonary arterial hypertension	Thrombospondin-1 and integrin-b3 may increase PASMC migration and proliferation
Ref (24)	Das et al.	Cancer cells	MMPs control integrin signalling. Gross MMP inhibition in cancer cell lines lowers traction pressures, cell rounding, and cell softness through integrin-b1 levels and localization.
Ref [26]	Ando et al.	Cancer cells	The TIMP-1- Yes-associated protein (YAP)/ transcriptional co-activator with PDZ binding motif (TAZ) axis may be a new cancer therapeutic target since it increases cell proliferation in cancer.
Ref (25)	Jia et al.	Ischemic Stroke	PAH development increases pulmonary vasculature MMP-8, a new protective factor. MMP-8 disrupts PASMC integrin- $\beta$ 3/FAK and YAP/TAZ-dependent mechanical signalling and matrix composition to resist pathologic mechanobiological feedback
Ref (26)	Wu et al.	Mice	Neuropeptide Y upregulates MMP-8, which increases macrophage migration and neointima development.
Ref (36)	Iwańczyk et al.	coronary artery abnormal dilatation patients	MMP-8 to aberrant coronary artery dilation
Ref (37)	Zhang et al.	Mice	Increased MMP-8 levels in aortic dissection patients
Ref (38)	Maghajothi et al.	Hypertensive	MMP-8 promoter GTT haplotype reduces NF-B-mediated MMP-8 production during inflammation and increases hypertension risk
Ref (39)	Cárcel-Márquez et al.	Ischemic Stroke	Lower serum MMP-12 levels to ischemic stroke, lower MMP-1 and MMP-12 to large-artery stroke, and higher MMP-8 to lacunar stroke.
Ref (40)	Yang et al.	Adventitia stem/progenitor cells	Macrophage-derived MMP8 modulates AdSPC-to-SMC differentiation and injury-induced neointimal SMC hyperplasia. The results on MMP8's role in AdSPC differentiation and neointima development in angiographic restenosis may help develop new medications to prevent it.
Ref (41)	Mitrut et al.	Dilated cardiomyopathy	TIMP-MMP imbalance disturbs ECM architecture and leads to dilated cardiomyopathy (DCM) remodelling, which may be utilised to design novel therapeutic treatments.
Ref (42)	Hashimoto et al.	Aortic dissection	In a rare aortic rupture caused by MSSA pneumonia, neutrophil proteases, including MMPs, were implicated

OPN: Osteopontin, PAH: Pulmonary Arterial Hypertension (Pulmoner Arteriyel Hipertansiyon), TGF- $\beta$ : Transforming Growth Factor Beta, CD47: Cluster of Differentiation 47, cMyc: Cellular Myc (a proto-oncogene), TSP1: Thrombospondin-1, PASMC: Pulmonary Artery Smooth Muscle Cell, MMP: Matrix Metalloproteinase, TIMP: Tissue Inhibitor of Metalloproteinases, YAP: Yes-associated Protein, TAZ: Transcriptional Co-activator with PDZ-binding Motif, FAK: Focal Adhesion Kinase, NF- $\kappa$ B: Nuclear Factor Kappa-light-chain-enhancer of Activated B Cells, AdSPC: Adventitial Stem/Progenitor Cell, SMC: Smooth Muscle Cell, ECM: Extracellular Matrix, DCM: Dilated Cardiomyopathy, MSSA: Methicillin-Sensitive Staphylococcus Aureus.

fibrous caps, weakening them and increasing the risk of plaque rupture, which may cause a heart attack or stroke. Heart failure impairs cardiac function due to MMP-8-induced ventricular remodelling. MMP-8 and CVDs are tricky. The equilibrium between MMP-8 and its tissue inhibitors, TIMPs, determines MMP-8's effect on CVDs. MMP-8 inhibitors may minimize CVD matrix breakdown and tissue injury. We need more research on MMP-8 and CVDs. Medical professionals should diagnose and treat CVDs and other illnesses.

## DECLERATIONS

**Ethics Committee Approval:** Since this study utilized publicly available literature, approval from the institutional review board was not obtained.

**Financial Disclosure:** The authors declared that this study has received no financial support.

**Author Contributions:** All of the authors declare that they have all participated in the design, execution, and analysis of the paper, and that they have approved the final version

**Conflict of Interest Statement:** The authors have no conflicts of interest to declare.

**Informed consent form:** Not Available

**Funding source:** No funding was received for the research

**Artificial Intelligence:** Artificial intelligence is utilized in this paper.

## REFERENCES

- Lenglet S, Mach F, Montecucco F. Role of matrix metalloproteinase-8 in atherosclerosis. *Mediators Inflamm.* 2013;2013:659282. doi:10.1155/2013/659282
- Ye S. Putative targeting of matrix metalloproteinase-8 in atherosclerosis. *Pharmacol Ther.* 2015;147:111-22.
- Ye S. Putative targeting of matrix metalloproteinase-8 in atherosclerosis. *Pharmacol Ther.* 2015;147:111-122. doi:10.1016/j.pharmthera.2014.11.007
- Wilson WR, Schwalbe EC, Jones JL, Bell PR, Thompson MM. Matrix metalloproteinase 8 (neutrophil collagenase) in the pathogenesis of abdominal aortic aneurysm. *Br J Surg.* 2005;92(7):828-833. doi:10.1002/bjs.4993
- Turu MM, Krupinski J, Catena E, et al. Intraplaque MMP-8 levels are increased in asymptomatic patients with carotid plaque progression on ultrasound. *Atherosclerosis.* 2006;187(1):161-169. doi:10.1016/j.atherosclerosis.2005.08.039
- Magyar MT, Szikszai Z, Balla J, et al. Early-onset carotid atherosclerosis is associated with increased intima-media thickness and elevated serum levels of inflammatory markers. *Stroke.* 2003;34(1):58-63. doi:10.1161/01.str.0000048845.83285.ac
- Raitio A, Tuomas H, Kokkonen N, et al. Levels of matrix metalloproteinase-2, -9 and -8 in the skin, serum and saliva of smokers and non-smokers. *Arch Dermatol Res.* 2005;297(6):242-248. doi:10.1007/s00403-005-0597-1
- Desvarieux M, Demmer RT, Rundek T, et al. Periodontal microbiota and carotid intima-media thickness: the Oral Infections and Vascular

- Disease Epidemiology Study (INVEST). *Circulation*. 2005;111(5):576-582. doi:10.1161/01.CIR.0000154582.37101.15
8. Tuomainen AM, Nyyssönen K, Laukkanen JA, et al. Serum matrix metalloproteinase-8 concentrations are associated with cardiovascular outcome in men. *Arterioscler Thromb Vasc Biol*. 2007;27(12):2722-2728. doi:10.1161/ATVBAHA.107.154831
  9. Kato R, Momiyama Y, Ohmori R, Taniguchi H, Nakamura H, Ohsuzu F. Plasma matrix metalloproteinase-8 concentrations are associated with the presence and severity of coronary artery disease. *Circ J*. 2005;69(9):1035-1040. doi:10.1253/circj.69.1035
  10. Kato R, Momiyama Y, Ohmori R, Taniguchi H, Nakamura H, Ohsuzu F. Levels of matrix metalloproteinase-1 in patients with and without coronary artery disease and relation to complex and noncomplex coronary plaques. *Am J Cardiol*. 2005;95(1):90-92. doi:10.1016/j.amjcard.2004.08.066
  11. Lim SY, Jeong MH, Bae EH, et al. Predictive factors of major adverse cardiac events in acute myocardial infarction patients complicated by cardiogenic shock undergoing primary percutaneous coronary intervention. *Circ J*. 2005;69(2):154-158. doi:10.1253/circj.69.154
  12. Death AK, Fisher EJ, McGrath KC, Yue DK. High glucose alters matrix metalloproteinase expression in two key vascular cells: potential impact on atherosclerosis in diabetes. *Atherosclerosis*. 2003;168(2):263-269. doi:10.1016/s0021-9150(03)00140-0
  13. Pullamsetti S, Krick S, Yilmaz H, et al. Inhaled tolfenetrine reverses pulmonary vascular remodeling via inhibition of smooth muscle cell migration. *Respir Res*. 2005;6(1):128. Published 2005 Nov 1. doi:10.1186/1465-9921-6-128
  14. Araujo JA, Zhang M, Yin F. Heme oxygenase-1, oxidation, inflammation, and atherosclerosis. *Front Pharmacol*. 2012;3:119. Published 2012 Jul 19. doi:10.3389/fphar.2012.00119
  15. Rosenberg M, Meyer FJ, Gruenig E, et al. Osteopontin predicts adverse right ventricular remodeling and dysfunction in pulmonary hypertension. *Eur J Clin Invest*. 2012;42(9):933-942. doi:10.1111/j.1365-2362.2012.02671.x
  16. Mura M, Cecchini MJ, Joseph M, Granton JT. Osteopontin lung gene expression is a marker of disease severity in pulmonary arterial hypertension. *Respirology*. 2019;24(11):1104-1110. doi:10.1111/resp.13557
  17. Saker M, Lipskaia L, Marcos E, et al. *Osteopontin, a Key Mediator Expressed by Senescent Pulmonary Vascular Cells in Pulmonary Hypertension*. *Arterioscler Thromb Vasc Biol*. 2016;36(9):1879-1890. doi:10.1161/ATVBAHA.116.307839
  18. Anwar A, Li M, Frid MG, et al. Osteopontin is an endogenous modulator of the constitutively activated phenotype of pulmonary adventitial fibroblasts in hypoxic pulmonary hypertension. *Am J Physiol Lung Cell Mol Physiol*. 2012;303(1):L1-L11. doi:10.1152/ajplung.00050.2012
  19. Kumar R, Mickael C, Kassa B, et al. TGF- $\beta$  activation by bone marrow-derived thrombospondin-1 causes Schistosoma- and hypoxia-induced pulmonary hypertension. *Nat Commun*. 2017;8:15494. Published 2017 May 30. doi:10.1038/ncomms15494
  20. Kaiser R, Frantz C, Bals R, Wilkens H. The role of circulating thrombospondin-1 in patients with precapillary pulmonary hypertension. *Respir Res*. 2016;17(1):96. Published 2016 Jul 30. doi:10.1186/s12931-016-0412-x
  21. Rogers NM, Yao M, Sembrat J, et al. Cellular, pharmacological, and biophysical evaluation of explanted lungs from a patient with sickle cell disease and severe pulmonary arterial hypertension. *Pulm Circ*. 2013;3(4):936-951. doi:10.1086/674754
  22. Rogers NM, Sharifi-Sanjani M, Yao M, et al. TSP1-CD47 signaling is upregulated in clinical pulmonary hypertension and contributes to pulmonary arterial vasculopathy and dysfunction. *Cardiovasc Res*. 2017;113(1):15-29. doi:10.1093/cvr/cvw218
  23. Rogers NM, Ghimire K, Calzada MJ, Isenberg JS. Matricellular protein thrombospondin-1 in pulmonary hypertension: multiple pathways to disease. *Cardiovasc Res*. 2017;113(8):858-868. doi:10.1093/cvr/cvx094
  24. Das A, Monteiro M, Barai A, Kumar S, Sen S. MMP proteolytic activity regulates cancer invasiveness by modulating integrins. *Sci Rep*. 2017;7(1):14219. Published 2017 Oct 27. doi:10.1038/s41598-017-14340-w
  25. Jung KK, Liu XW, Chirco R, Fridman R, Kim HR. Identification of CD63 as a tissue inhibitor of metalloproteinase-1 interacting cell surface protein. *EMBO J*. 2006;25(17):3934-3942. doi:10.1038/sj.emboj.7601281
  26. Ando T, Charindra D, Shrestha M, et al. Tissue inhibitor of metalloproteinase-1 promotes cell proliferation through YAP/TAZ activation in cancer. *Oncogene*. 2018;37(2):263-270. doi:10.1038/onc.2017.321
  27. Pellinen T, Rantala JK, Arjonen A, Mpindi JP, Kallioniemi O, Ivaska J. A functional genetic screen reveals new regulators of  $\beta$ 1-integrin activity. *J Cell Sci*. 2012;125(Pt 3):649-661. doi:10.1242/jcs.090704
  28. Nardone G, Oliver-De La Cruz J, Vrbsky J, et al. YAP regulates cell mechanics by controlling focal adhesion assembly. *Nat Commun*. 2017;8:15321. Published 2017 May 15. doi:10.1038/ncomms15321
  29. Varadarajulu J, Laser M, Hupp M, Wu R, Hauck CR. Targeting of alpha(v) integrins interferes with FAK activation and smooth muscle cell migration and invasion. *Biochem Biophys Res Commun*. 2005;331(2):404-412. doi:10.1016/j.bbrc.2005.03.175
  30. Ron A, Azeloglu EU, Calizo RC, et al. Cell shape information is transduced through tension-independent mechanisms. *Nat Commun*. 2017;8(1):2145. Published 2017 Dec 15. doi:10.1038/s41467-017-02218-4
  31. Jia Y, Guo D, Zhang K, et al. Causal associations of serum matrix metalloproteinase-8 level with ischaemic stroke and ischaemic stroke subtypes: a Mendelian randomization study. *Eur J Neurol*. 2021;28(8):2543-2551. doi:10.1111/ene.14878
  32. Dieffenbach PB, Mallarino Haeger C, Rehman R, et al. A Novel Protective Role for Matrix Metalloproteinase-8 in the Pulmonary Vasculature. *Am J Respir Crit Care Med*. 2021;204(12):1433-1451. doi:10.1164/rccm.202108-1863OC
  33. Cuadrado E, Rosell A, Penalba A, et al. Vascular MMP-9/TIMP-2 and neuronal MMP-10 up-regulation in human brain after stroke: a combined laser microdissection and protein array study. *J Proteome Res*. 2009;8(6):3191-3197. doi:10.1021/pr801012x
  34. Lorenz S, De Pasquale G, Segal AZ, Beal MF. Dysregulation of the levels of matrix metalloproteinases and tissue inhibitors of matrix metalloproteinases in the early phase of cerebral ischemia. *Stroke*. 2003;34(6):e37-e38. doi:10.1161/01.STR.0000075563.45920.24
  35. Wu W, Peng S, Shi Y, Li L, Song Z, Lin S. NPY promotes macrophage migration by upregulating matrix metalloproteinase-8 expression. *J Cell Physiol*. 2021;236(3):1903-1912. doi:10.1002/jcp.29973
  36. Iwańczyk S, Lehmann T, Grygier M, Woźniak P, Lesiak M, Araszkiewicz A. Serum matrix metalloproteinase-8 level in patients with coronary artery abnormal dilatation. *Pol Arch Intern Med*. 2022;132(5):16241. doi:10.20452/pamw.16241
  37. Zhang C, Niu K, Ren M, et al. Targeted Inhibition of Matrix Metalloproteinase-8 Prevents Aortic Dissection in a Murine Model. *Cells*. 2022;11(20):3218. Published 2022 Oct 14. doi:10.3390/cells11203218
  38. Maghajothe S, Subramanian L, Mani P, et al. A common Matrix metalloproteinase 8 promoter haplotype enhances the risk for hypertension via diminished interactions with nuclear factor kappa B. *J Hypertens*. 2022;40(11):2147-2160. doi:10.1097/HJH.0000000000003234
  39. Cárcel-Márquez J, Culléll N, Muñio E, et al. Causal Effect of MMP-1 (Matrix Metalloproteinase-1), MMP-8, and MMP-12 Levels on Ischemic Stroke: A Mendelian Randomization Study. *Stroke*. 2021;52(7):e316-e320. doi:10.1161/STROKEAHA.120.033041
  40. Yang F, Chen Q, Yang M, et al. Macrophage-derived MMP-8 determines smooth muscle cell differentiation from adventitia stem/progenitor cells and promotes neointima hyperplasia. *Cardiovasc Res*. 2020;116(1):211-225. doi:10.1093/cvr/cvz044
  41. Mitrut R, Stepan AE, Mărgărețescu C, et al. Immunoeexpression of MMP-8, MMP-9 and TIMP-2 in dilated cardiomyopathy. *Rom J Morphol Embryol*. 2019;60(1):119-124.
  42. Hashimoto M, Kuriwa S, Kojima A, et al. Aortic rupture involving matrix metalloproteinases 8 and 9 during Staphylococcus aureus pneumonia. *Thorax*. 2018;73(4):397-398. doi:10.1136/thoraxjnl-2017-210784